

The shift from control to elimination: implications for diagnostics

Fabian Schär, PhD

January 31, 2017

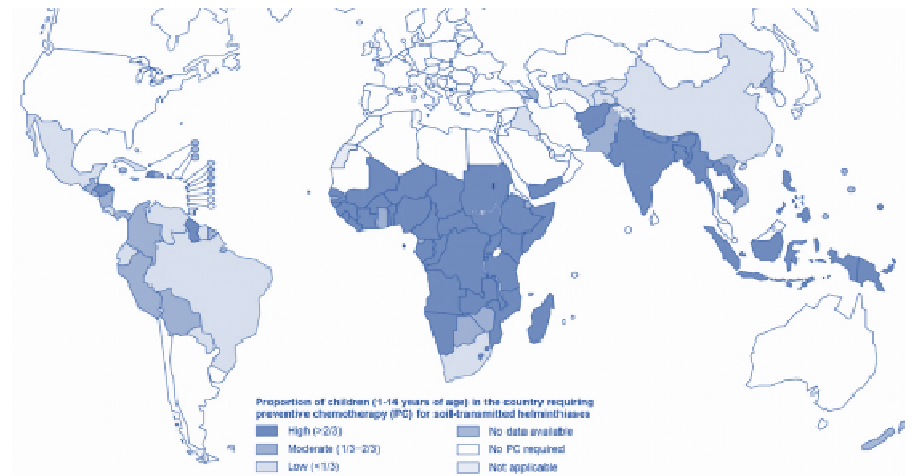
LCNTDR

deworm³

N **NATURAL
HISTORY
MUSEUM**

Introduction

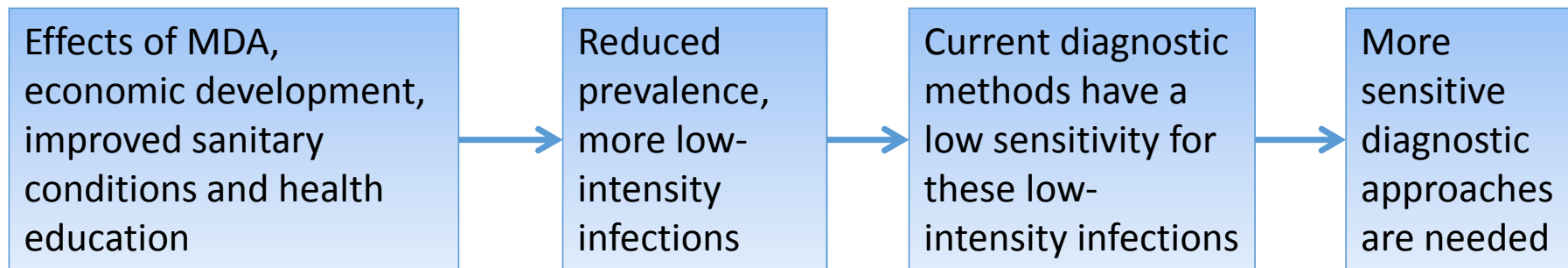
- Soil-transmitted helminths (STHs) are part of the NTDs and account for a high burden of disease
- Morbidity control with MDA is the current STH strategy
- Future strategies should aim at moving from control to elimination of STHs



World Health Organization (WHO) 2014

The challenge

The switch from control to elimination from the diagnostic point of view



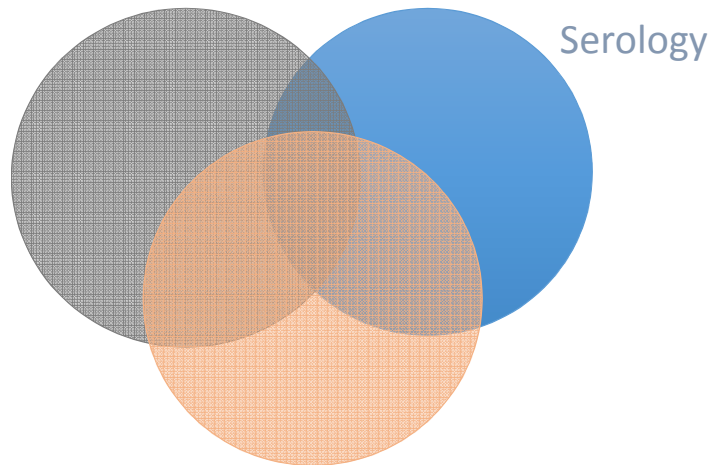
Importance of detecting low-intensity infections

Current and new diagnostic methods

	Coprological stool methods	Molecular methods
Advantages	<ul style="list-style-type: none">• Cheap• Readily available• Easy to apply in field settings	<ul style="list-style-type: none">• Sensitivity drastically increased
Disadvantages	<ul style="list-style-type: none">• Lower sensitivity• Risk of missing low intensity infections	<ul style="list-style-type: none">• Higher costs• Difficult to implement in field settings• Risk of contamination

Validation of diagnostic methods

Molecular
Methods



Coprological
methods

Adapted from Johansen; Acta Tropica 141
(2015) 161-169

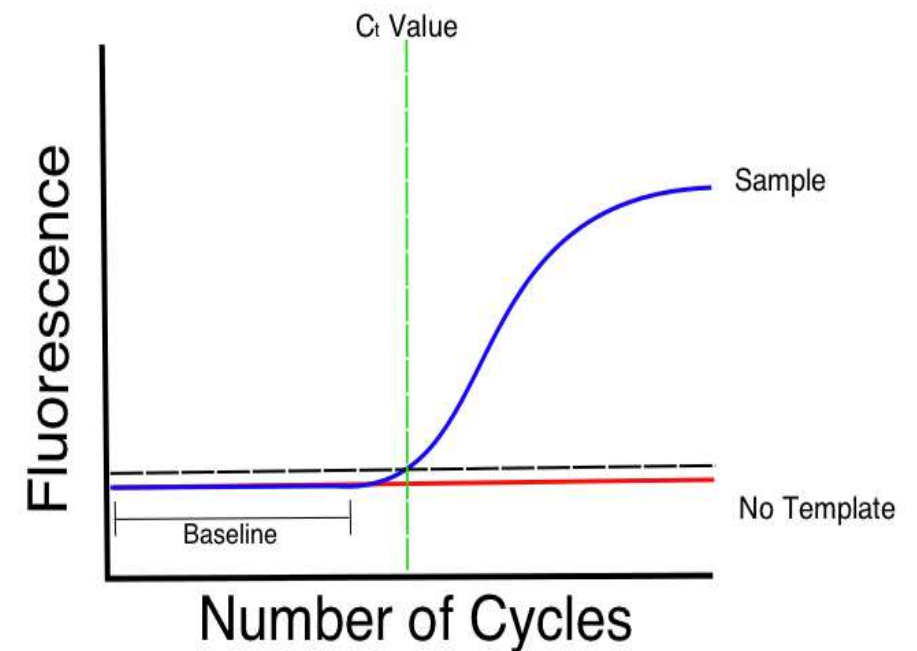
The lack of gold standard
diagnostic methods makes
validation of new diagnostic
approaches challenging

Advances in PCR methodology

- Ribosomal and mitochondrial targets in PCR may be sub-optimal for low-intensity infections and lack of species-specificity
- New PCR approaches have been developed that target species-specific repetitive non-coding repeat DNA
- The high repetition sequences have a rapid evolutionary divergence: ideal for distinguishing even closely related species (i.e. *A. duodenale* and *A. ceylanicum*)
- Molecular approaches are becoming cheaper, easier to conduct and field based methods are in development
- PCR enables to detect smallest volumes of parasite DNA, ideal for detecting low-intensity infections.

Pilotte et al (2016) PLoS Negl Trop Dis 10(3)

de**worm**³



Quantitative PCR (qPCR)

- qPCR allows to **quantify** the detected genetic material (DNA, RNA)
- qPCR is faster in detecting amplified DNA, no separate readout is needed
- Sensitivity is increased
- Lower amounts of material can be used
- Throughput is considerably higher than conventional PCR
- **Possible challenges: qPCR is more expensive and more difficult to implement in field settings**
- Today's possibility of multi-parallel assays helps to drastically reduce cost of the PCR
- Additionally, the assay can be optimized for the geographic location (i.e. what pathogens should be included) as the assays can run independently
- Pooling of samples can further reduce costs

Llewellyn et al (2016). PLoS Negl Trop Dis 10(1): Gordon et al (2015). Int Jnl for Parasitology 45 (2015) 477–483

qPCR in DeWorm3

- Define molecular cut-offs to define transmission interruption
- Optimize stool collection, processing and storage
- Identify optimal extraction and pooling methodologies
- Build capacity at multiple sites for a systematic and harmonized approach to molecular diagnosis of STHs
- Create a biobank of stored parasite genetic network for the NTD community

deworm³

deworm³

Acknowledgments

Dr. Judd Walson, PI
Iain Gardiner
Arianna Rubin Means
Dr. Kristjana Ásbjörnsdóttir
Dr. Fabian Schär
Anoushka Bassett
Elodie Yard
Leanne Doran
Dr. Bryan Weiner
Dr. Adam Szpiro

Prof. Gagandeep Kang
Prof. Robin Bailey
Dr. Khumbo Kalua
Dr. Adrian Luty
Dr. Mouda Ibikounlé
Dr. Rachel Pullan
Dr. Kate Halliday

Dr. Tim Littlewood
Dr. David Rollinson
Sir Roy Anderson
Dr. James Truscott
Dr. Marleene Werkman
Dr. Sam Farrell
Dr. James Wright



BILL & MELINDA
GATES *foundation*

LONDON
SCHOOL of
HYGIENE
& TROPICAL
MEDICINE



de**worm**³

Imperial College
London